

INTEGRATED GANODERMA MANAGEMENT IN OIL PALM PLANTATION IN MALAYSIA

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OIL PALM INDUSTRY IN MALAYSIA



• The oil palm is an important commodity in Southeast Asia (SEA).

 Oil palm is the major commodity crop for Malaysia and the fourth-largest contributor to Malaysia's national revenue.

• The total area of oil palm in Malaysia is 5.6 million hectares, with a total export revenue of palm oil and palm-based products valued of more than USD 20 billion in 2024.

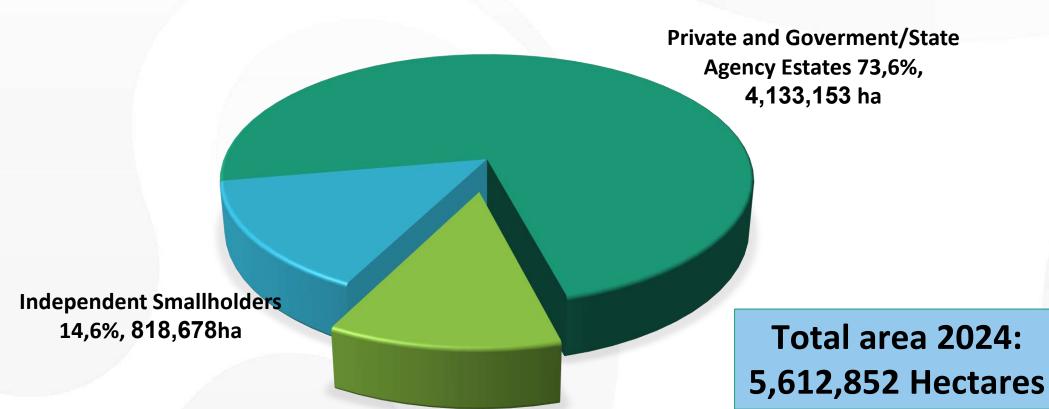




OIL PALM INDUSTRY IN MALAYSIA



MALAYSIA: OIL PALM PLANTED AREA BY CATEGORY, 2024



Organised Smallholders 11,8%, 661,021 ha

OIL PALM INDUSTRY IN MALAYSIA



MALAYSIAN PALM OIL EXPORT DESTINATIONS, 2024







1) INDIA

3.03 Mn T 1 6.5%





2) CHINA 1.39 Mn T \$ 5.3%







1.26 Mn T 1 37.7%





5) TURKIYE 0.91 Mn T 12.4%



6) PHILIPPINES 0.69 Mn T \$55.5%

1.29 Mn T 1 21.3%

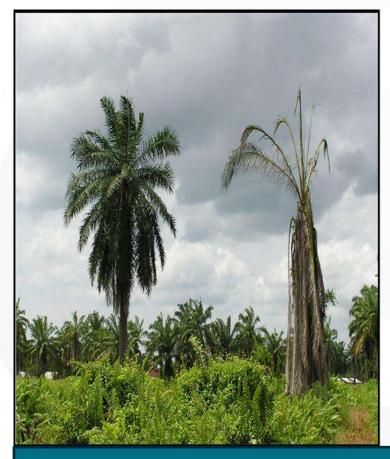
Source: MPOB



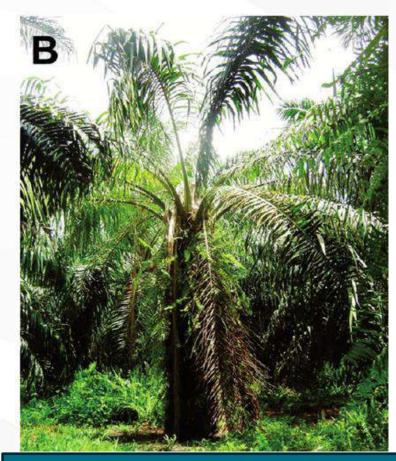
MALAYSIAN PALM OIL BOARD

MAJOR OIL PALM DISEASES





Basal Stem Rot Disease



Bud Rot Disease

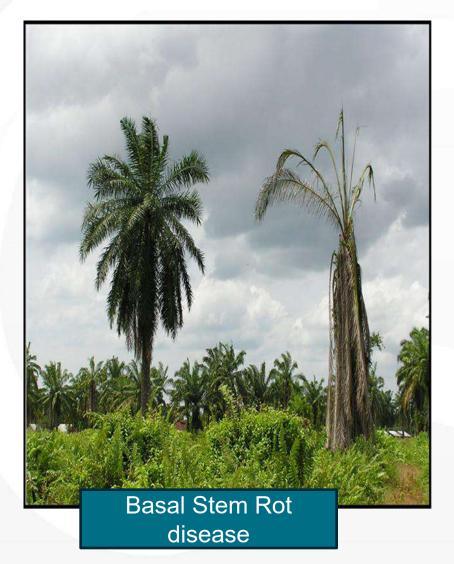


Vascular Wilt Disease

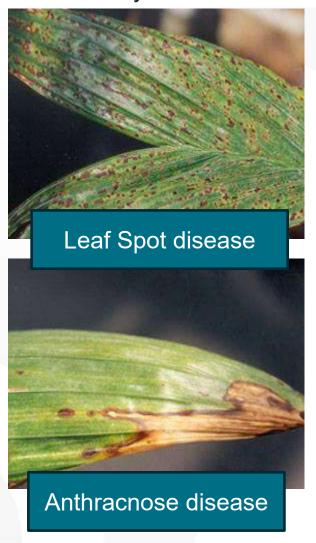
Exotic diseases

OIL PALM DISEASES IN MALAYSIA

Field diseases

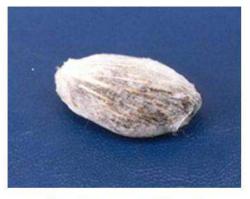


Nursery Diseases





Seed Diseases



Biji benih yang dijangkiti dilitupi dengan miselium putih



Biji benih dijangkiti dengan miselium putih dan bintik kehijauan (spora)

EMERGING DISEASES IN MALAYSIA



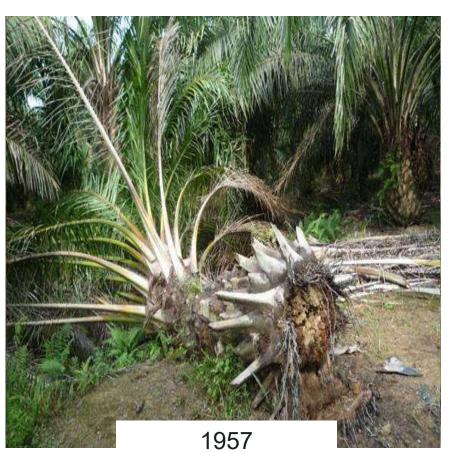


BASALS EN ROTD SEASE INTERNACIONAL SOBRE PALMA DE ACE



History of BSR disease incidences in Malaysia







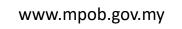
Old palm > 25 years old



Mature palms, 10 -15 years old



Immature palms, 1- 2 years old

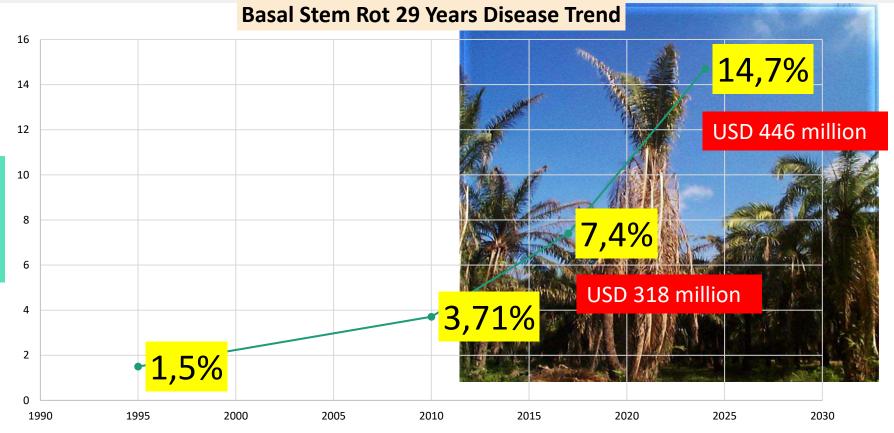




Updated status of Basal Stem Rot (BSR) disease incidence in oil palm plantations in Malaysia

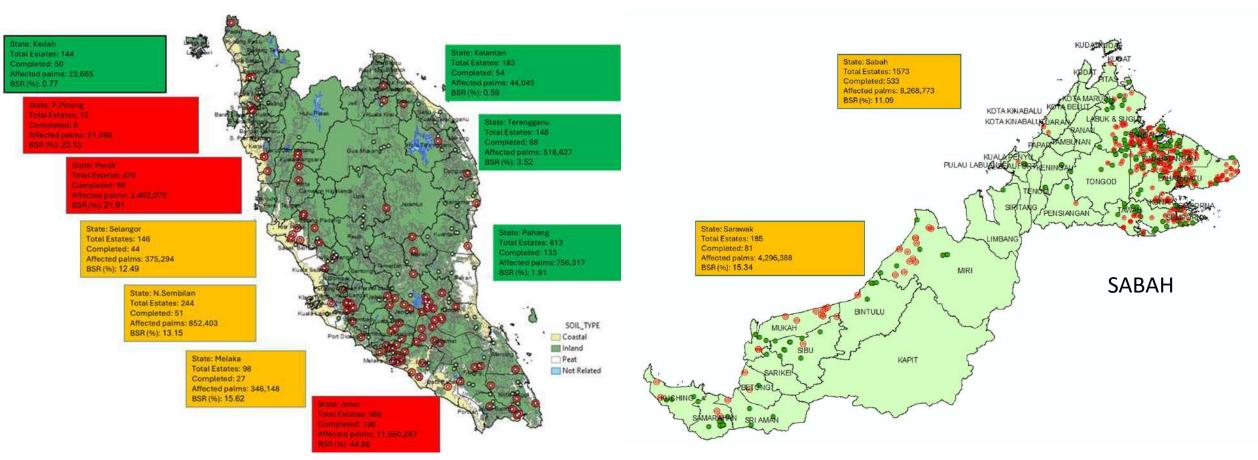
199, 633 hectares affected

29,545,684 trees infected



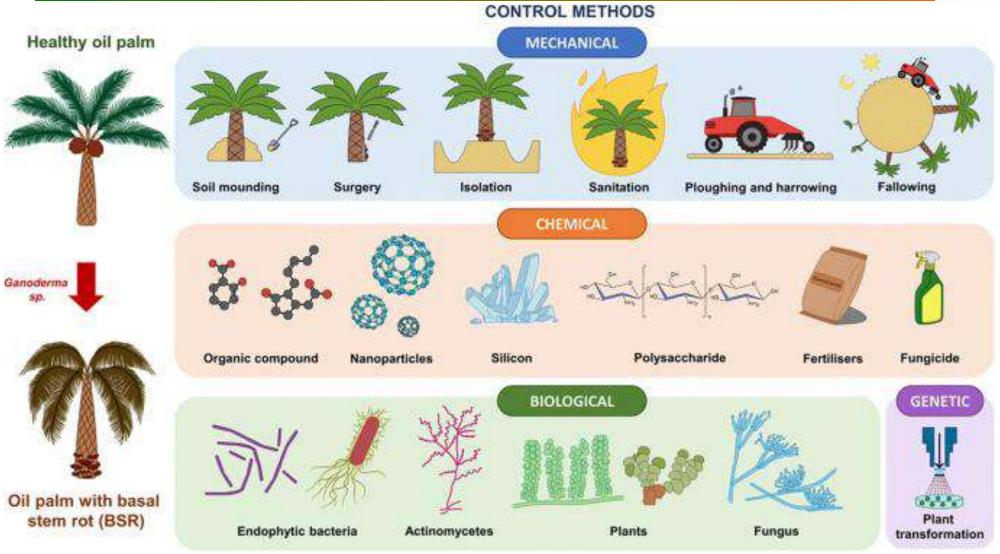


PENINSULAR MALAYSIA



SARAWAK

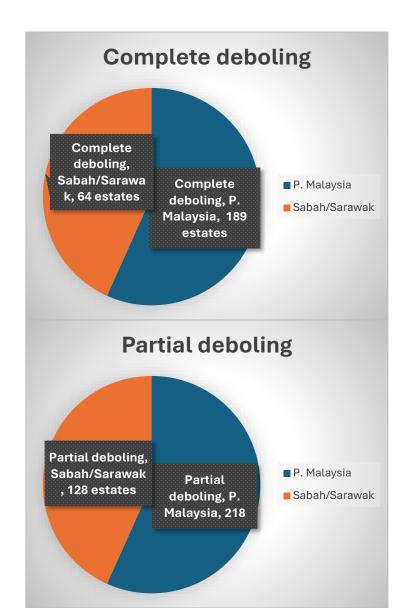






BASA STEWROLD SEASE INTERNACIONAL SOBRE PALMA DE ACEITE 21º International Oil Palm Conference 21º Internatio





Complete deboling

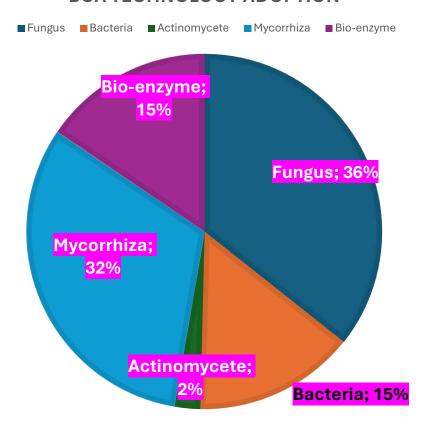
	Estates with	BSR disease	Treated area involved		
Region	No. of estates	Area (ha)	No. of estates	Area (ha)	
P. Malaysia	325	114,733.27	189	5,738.13	
Sabah/Sarawak	435	84,899.74	64	4,377.87	
Total	760	199,633.01	253	10,116.00	
Percentage (%)	54.7	13.7		5.07	

Partial deboling

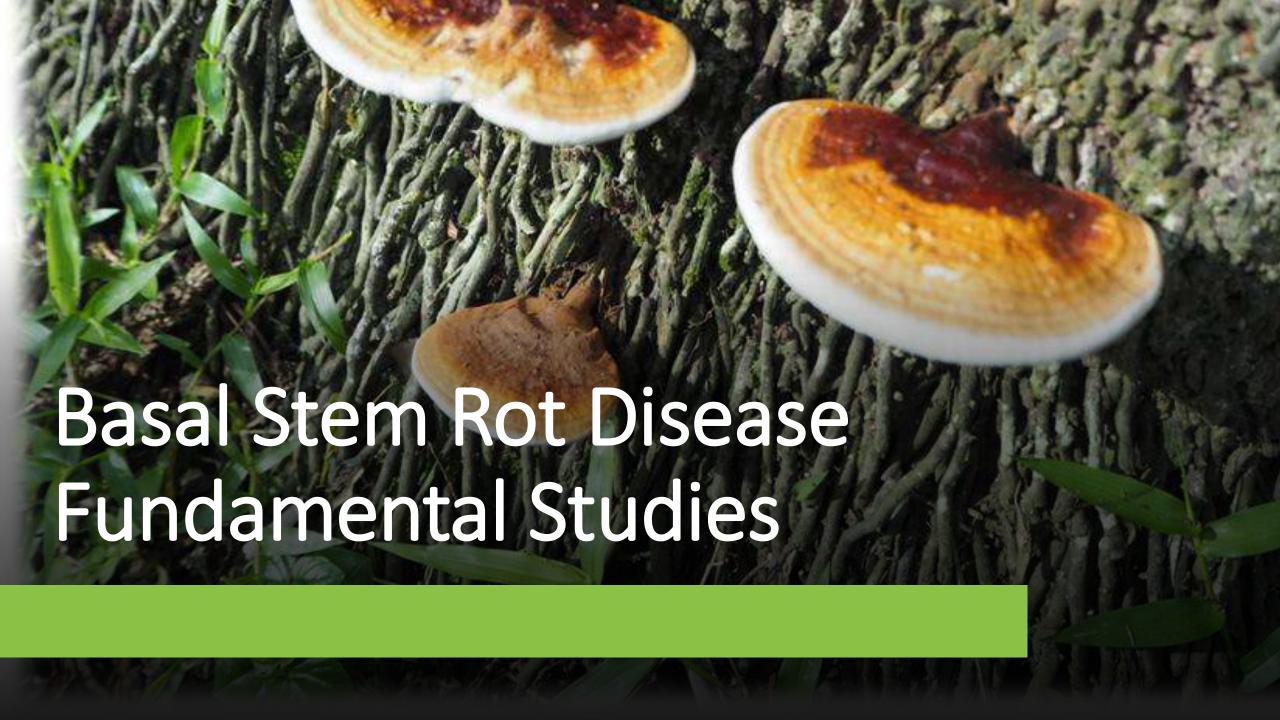
	Estates with	BSR disease	Treated area involved		
Region	No. of estates	Area (ha)	No. of estates	Area (ha)	
P. Malaysia	325	114,733.27	218	1,442.30	
Sabah/Sarawak	435	84,899.74	128	1,100.40	
Total	760	199,633.01	332	2,542.70	
Percentage (%)	54.7	13.7		1.27	



BCA TECHNOLOGY ADOPTION

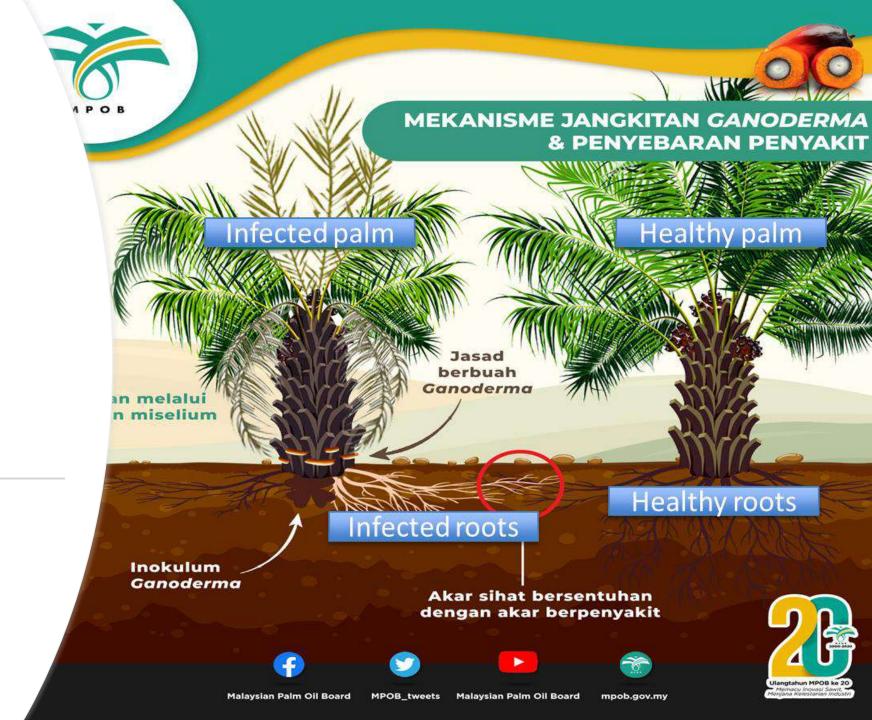


	Estates with BSR disease		Treated area involved		
Region	No. of estates	Area (ha)	No. of estates	Area (ha)	
P. Malaysia	325	114,733.27	95	4,303.7	
Sabah/Sarawak	435	84,899.74	34	1,396.0	
Total	760	199,633.01	129	5,699.8	





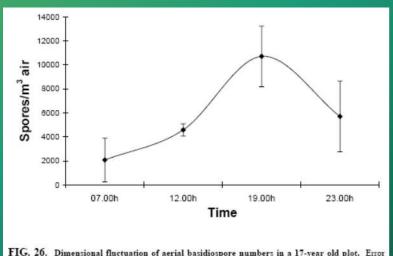
Mechanism of infection (Root To Root)



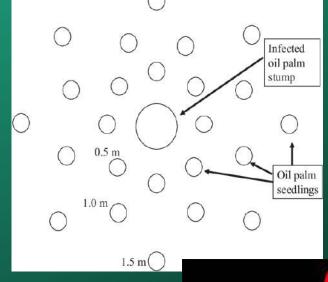


Mechanism of infection (Basidiospore)



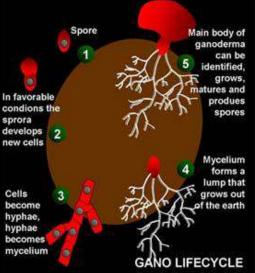


bars represent standard deviation of the mean of four samples at each time point



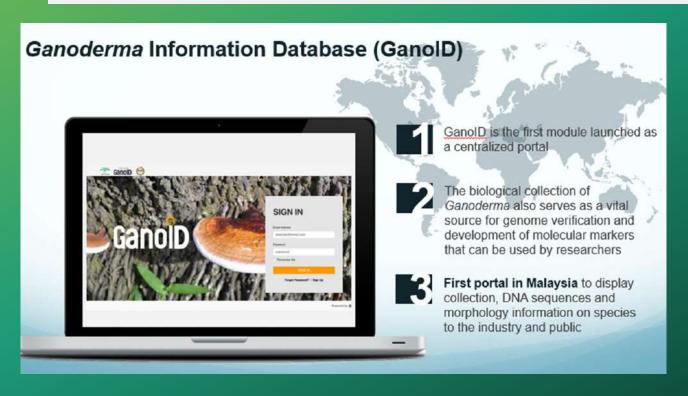
c. 10⁸ **spores** *per bracket* **per day** (Paul Bridge CABI)

- Infection through basidiospores (wind, insect vector etc.).
- Basidiospore stay active for 10 days.





Updataing on Ganoderma systematics: Diversity based on morphotaxonomy, ultrastructural, biochemical and molecular techniques













Ganoderma boninense is now the dominant species in Malaysia

Unique basidiocarps in different colours; sizes; Basidiospores ellipsoid



Effect of pH on Ganoderma boninense



- ☐ Impact in the field
- Low pH equals to low microbial population
- Low beneficial microbes to compete with *G. boninense*

- Grow over a wide range of pH ranging pH 3.0 to 9.0.
- pH 5 is the optimum pH for the growth of mycelium for all *Ganoderma* isolates.

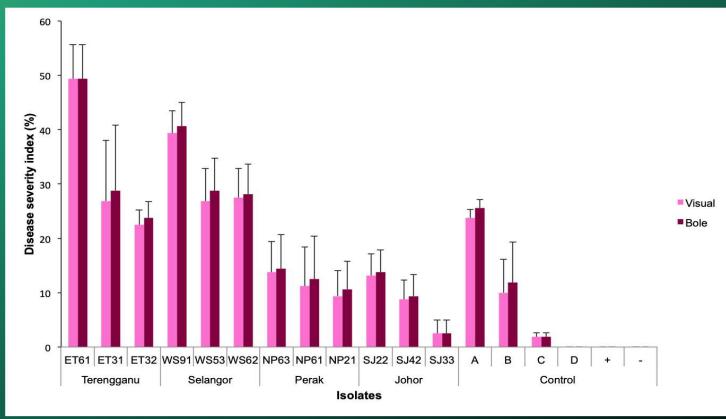


Differential Responses In Ganoderma Isolate Aggressiveness



COASTAL LAND





- Most aggressive : ET61 (Terengganu)
- Least aggressive : SJ 33 (Johor)
- Standard culture PER 71 recorded DSI of 23.8 %

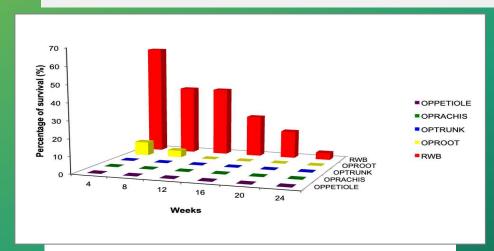


Differential Responses In Ganoderma Isolate Aggressiveness

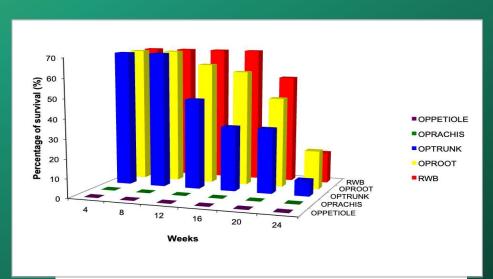
Disease Incidences (%)	Location	Soil Type	Palm symptom & Infection (%)	Palm (age)	Nursery DSI (%)
	Johor (SJ33)	Inland	< 25 %; Skirting	27	3.10
0 – 10	Perak (NP61)	Inland	< 25 %	27	13.80
0 10	Perak (NP21)	Inland	> 50 %	27	13.80
	Johor (SJ22)	Inland	< 25 % infected; Skirting	27	16.30
	Perak (NP63)	Inland	< 30 %; Skirting, multiple spears & hollow base	27	18.80
10 – 20	Terengganu (ET32)	Coastal	< 10 %; Asymptomatic	22 Iss	28.80
	Selangor (WS53)	Coastal	< 25 %; Hollow base	19	35.60
20 – 30	Terengganu (ET31)	Coastal	< 10 %; Asymptomatic	98 20 20	33.10
	Selangor (WS62)	Coastal	> 75 %; Skirting		38.00
	Selangor (WS91)	Coastal	> 75 %	<u>=</u> 19	43.80
30 – 40	Terengganu (ET61)	Coastal	> 75 %; Skirting, multiple spears	19	58.80



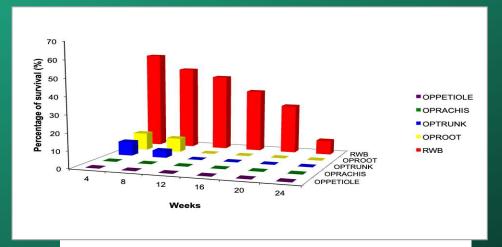
Survival of Ganoderma boninense In The Soil



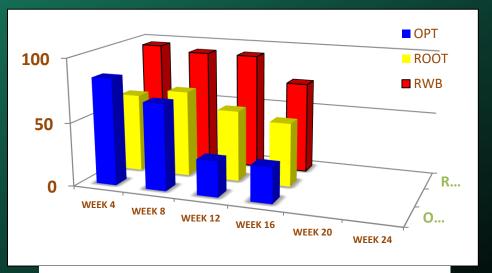
Recovery of *G. boninense* at 10 cm



Recovery of G. boninense at 30 cm



Recovery of G. boninense at 20 cm



Recovery of G. boninense at 60 cm



Survival of Ganoderma boninense In The Soil

Rubber wood block (RWB)

Oil palm roots

Oil palm trunk

Oil palm petiole rachis

- ✓ Depth significantly influences the survival of *G. boninense*.
- ✓ Rubber wood block inoculum found to be the highest surviving followed by root and oil palm trunk
- ✓ Oil palm petiole and rachis were not able to sustain the survival of *G. boninense* in soil



EFFECTS OF PLANTING DENSITY ON GANODERMA DISEASE INCIDENCE

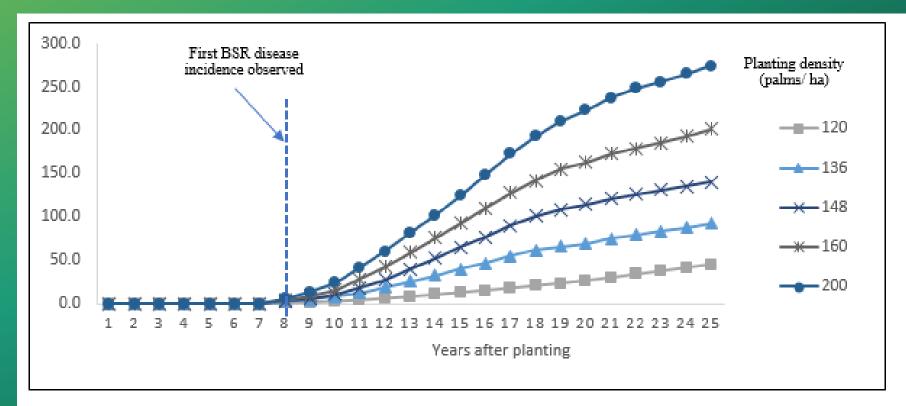


Figure 2. The BSR disease progress in different oil palm planting densities over the years after planting in the plantations.

Root to Root Contact?



Exploring the Effects of Herbicides on Bacterial Diversity and Functionality in Oil Palm Plantation Soils

1 Soil bacterial diversity were unaffected by herbicides

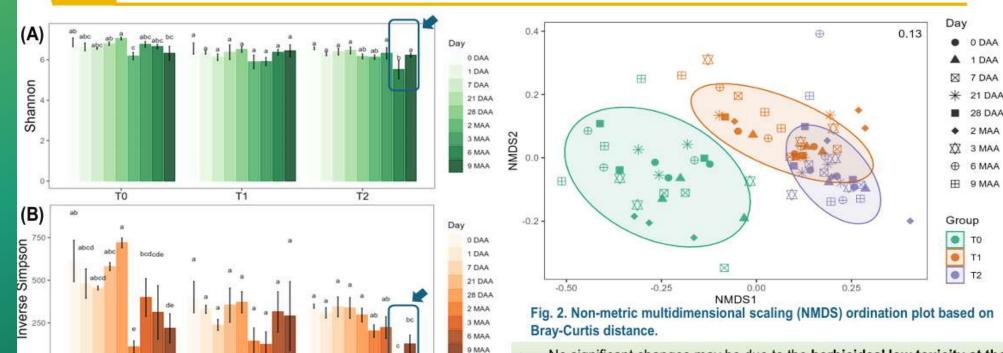


Fig. 1. Effects of herbicide application on Shannon (A) and inverse Simpson (B) diversity indices in control (T0), glufosinate-ammonium (T1), and metsulfuron-methyl (T2).

No significant changes may be due to the **herbicides' low toxicity at the** standard recommended rate⁵.

Vegetation loss from herbicide use affects soil microbes more than the chemicals themselves⁶. Metsulfuron-methyl spray drift can also alter plant species by reducing seed viability or plant vigour^{7,8}.

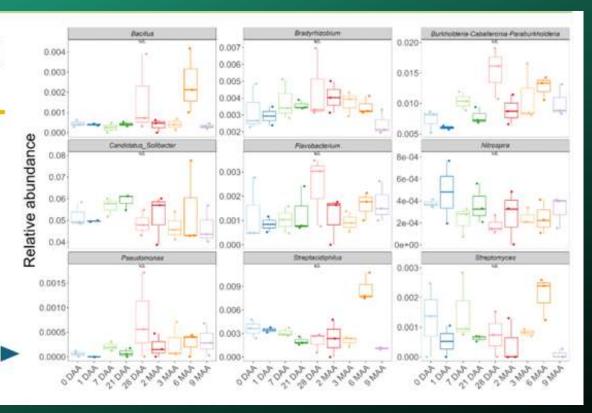


Exploring the Effects of Herbicides on Bacterial Diversity and Functionality in Oil Palm Plantation Soils

Beneficial soil rhizobacteria remained stable after application

- The acetolactate synthase (ALS) enzymes of beneficial bacteria differ in structure or regulatory mechanisms from those in plants⁹.
- Sulfonylurea herbicides like metsulfuron-methyl inhibit only type II and III
 acetohydroxyacid synthase (AHAS) isozymes in bacteria¹⁰.
- The abundance of beneficial bacteria did not change after spraying, probably due to genes coding for alternative AHAS isozymes¹¹.

Fig. 3. Relative abundance of key bacterial genera involved in soil health and plant growth in T2.





Exploring the Effects of Herbicides on Bacterial Diversity and Functionality in Oil Palm Plantation Soils

3 No significant impact on key soil functions and activities

- Cellulolytic bacteria that break down cellulose into oligosaccharides increased at 2 MAA. These bacterial groups prevent soil pollution and enhance fertility by maintaining organic carbon levels¹². Herbicides may suppress glucose-utilizing microorganisms, slowing cellulose conversion to glucose¹³.
- Bacterial groups related to ureolysis increased at 28
 DAA. Ureolytic bacteria utilise nitrogen for their urease activity, and since herbicides contain nitrogen, they may contribute to enhanced ureolysis in the soil¹⁴.
- Nitrification and nitrate reduction varied over the ninemonth study but showed no significant changes, likely due to functional gene redundancy in microbial communities¹⁵.

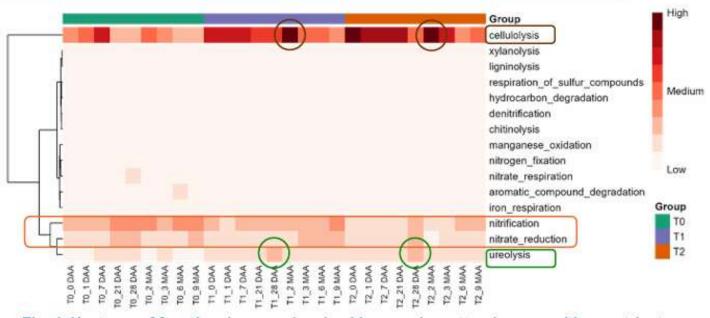
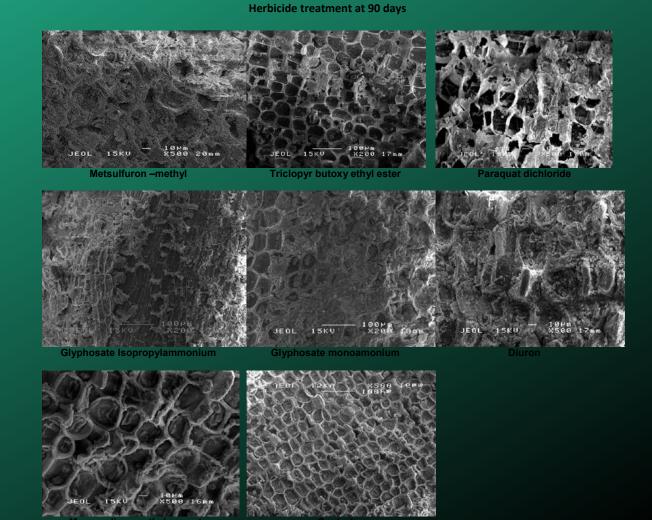


Fig. 4. Heatmap of functional groups involved in organic matter decomposition, nutrient cycling, and chemical degradation in soils applied with herbicides at various times.



Influence Of Herbicides On BSR Disease Incidence

- Any physical root injuries or any physiological disorders when the root samples were treated with the seven selected herbicides were observed.
- Plant roots treated with most of the herbicides showed large numbers of cell ruptures near the meristem of axes particularly plants treated with diuron (92.73% DI) followed by metsulfuron-methyl (83.27% DI) and glyphosate isopropylammonium (73.81% DI)

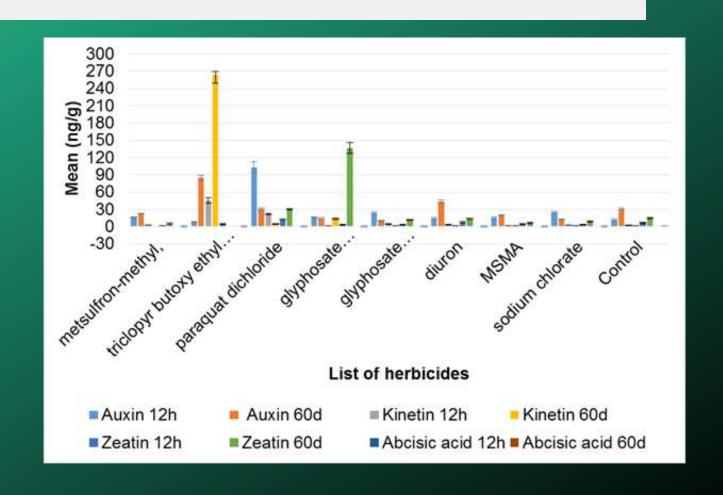




Influence Of Herbicides On BSR Disease Incidence

The root damages coincided with the reduction and imbalance of targeted phytohormomes (involved in plant growth regulation and plant defense system)

Plants treated with paraquat dichoride, results showed that the expression of auxin was reduced from 103.3 ng/g to 30.1 ng/g, kinetin (21.81 ng/g to 4 ng/g), zeatin (29.88 ng/g to 11.92 ng/g) and AA (0.14 ng/g to 0.081 ng/g)





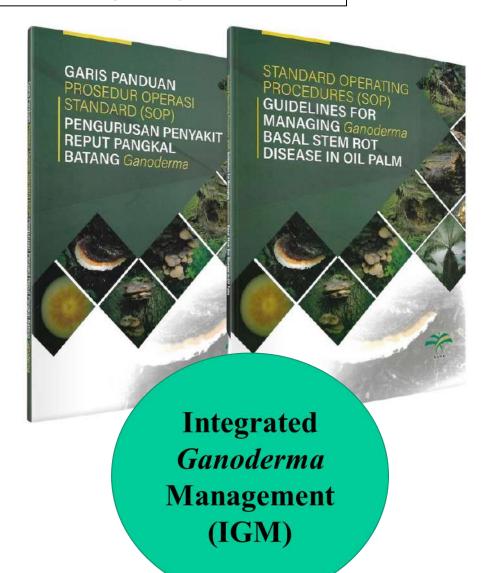
Management of Basal Stem Rot Disease



Standard Operating Procedure (SOP)

Over the years, MPOB continuously conducted research and advisory services related to Ganoderma disease with some of the technologies being eventually commercialised.

This standard operating procedure (SOP) for managing of Ganoderma disease in oil palm is based on the agreed standard practices as a guide for plantations to manage Ganoderma disease in oil palm more effectively.





Standard Operating Procedure (SOP)

Disease Census



Disease Symptoms Identification







Mature Palm

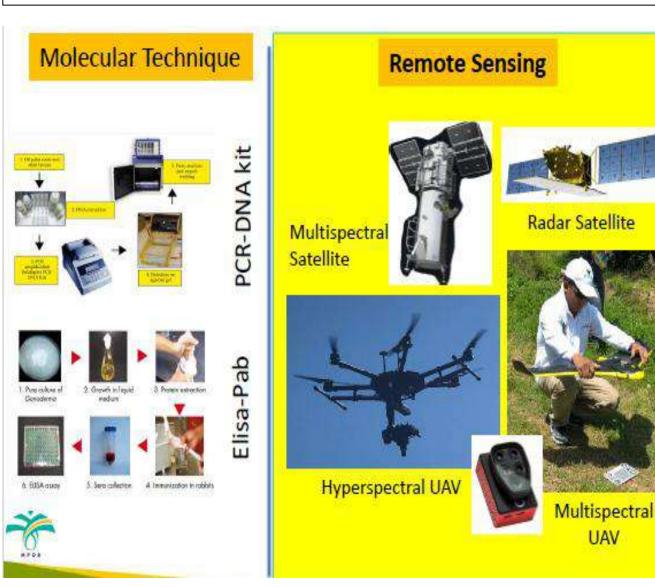
NOTE:

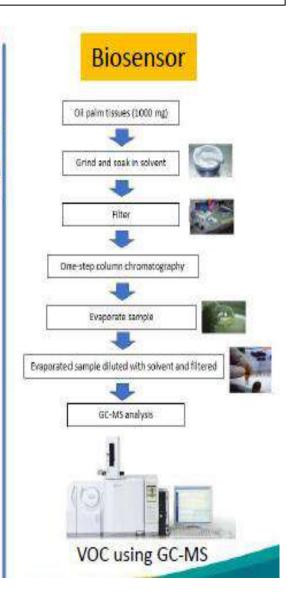
- A. BSR is a slow disease. Census can be conducted annually.
- B. Disease symptoms reference available on SawitSecure website

Early Disease Detection



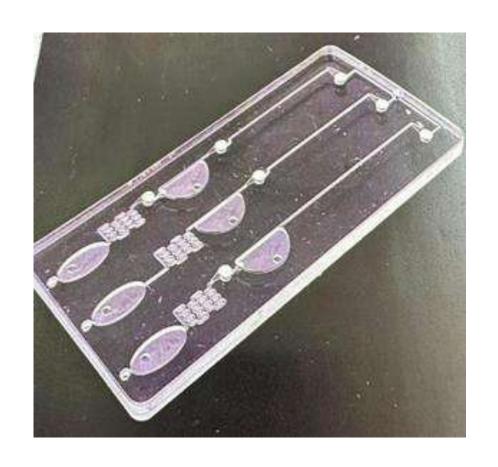
Standard Operating Procedure (SOP): Early Disease Detection

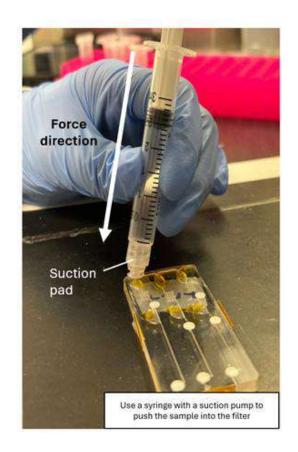


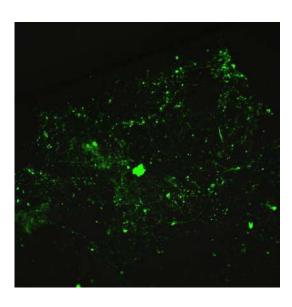




Standard Operating Procedure (SOP): Early Disease Detection







A diagnostic tool for *Ganoderma* detection (microfluidic nanotechnology)



Standard Operating Procedure (SOP): Existing Planting



Young/Immature palms

(< 10 years old)

Disease severity index (DSI)	Palm condition
DSI 0	Healthy palm
DSI 1	Mild, moderate, severely infected palm

SOP: Excavate and Replace



Mature palms

(> 10 years old)

Disease severity index (DSI)	Palm condition
DSI 0	Healthy palm
DSI 1	Early infection
DSI 2	Moderate infection
DSI 3	Severe infection
DSI 4	Dead

SOP: Manage Based on DSI

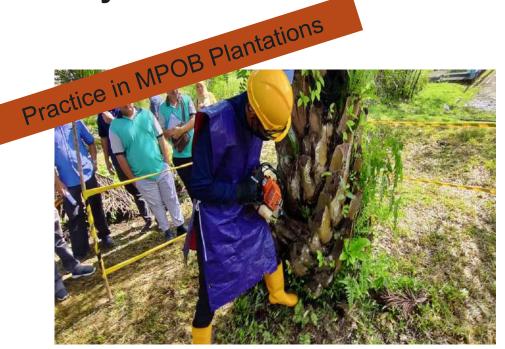


Standard Operating Procedure (SOP): Existing Planting

Focus on DSI 1 and DSI 2 only

Treatment with fungicide hexaconazole

- Prolong the economic life of an infected palm (fruiting).
- 66.6% 83.3% (average 74.4%, 3 trials) of treated infected palms with hexaconazole still alive and producing fruit bunches at 5 years and none from untreated palms.
- Cost: USD 14 per palm (2 treatments).
- In return: Average USD 125 in 5 years (Min).







Standard Operating Procedure (SOP): Existing Planting

Reference: Effect of Nanomaterials on Water and Solutes Translocation in Plants



Focus on DSI 1 and DSI 2 only

The technology allows a simultaneous nanonisation and encapsulation of hydrophobic crop protection inputs with a simple one step mixing process. This technology enables the inputs to acquire *phloem mobility*, enhanced *bioavailability* and *controlled release* properties.

Nano Adjuvant Hydrophobic actives Nano Adjuvant Hydrophobic actives Nano Adjuvant Hydrophobic actives Nano Adjuvant Nano Adjuvant Nano Adjuvant Nano Adjuvant Nano Adjuvant Nano Adjuvant Nanoparticles Nanoparticles

Hydroemission Nanotechnology for the Treatment of Ganoderma Basal Stem Rot

in Oil Palms







Standard Operating Procedure (SOP): Existing Planting

Focus on DSI 3 and DSI 4 only

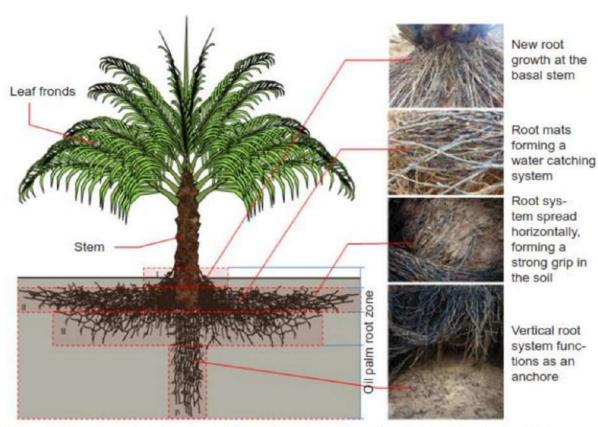


Figure 5 Oil palm roots architecture at the roots zone and its parts which is shown by the image resulted from the destructive measurement method.

Ganoderma boninense infects primary root

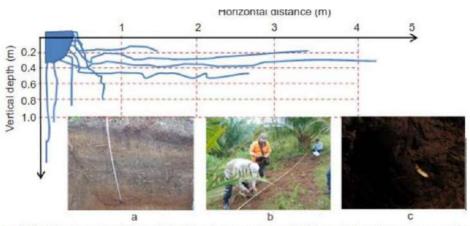


Figure 3 Architecture description of oil palm primary roots resulted from destructive measurement: a Excavation of soil around the palm roots; b Activity of measuring the root length of the palm at ground level; c The primary root tip that grows actively in the soil.



Standard Operating Procedure (SOP): Existing Planting

Focus on DSI 3 and DSI 4 only

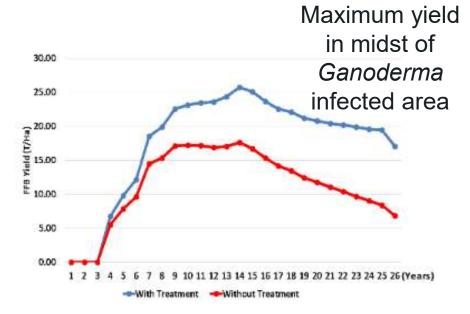
Removing the inocula pressure

- Sanitation / excavation (1.5m x 1.5m x 1.5m) to minimize the inoculum size.
- The excavated hole can be closed immediately with the nearby top soil.





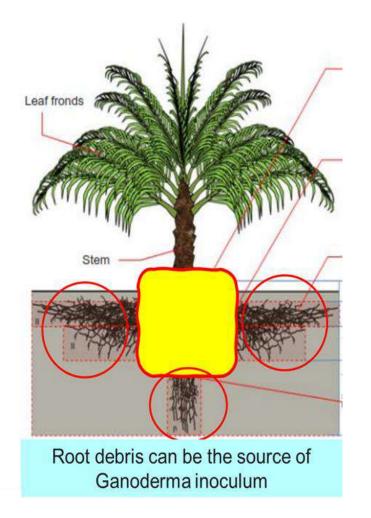
1.5m x 1.5m x 1.5m





Standard Operating Procedure (SOP): Existing Planting

Focus on DSI 3 and DSI 4 only



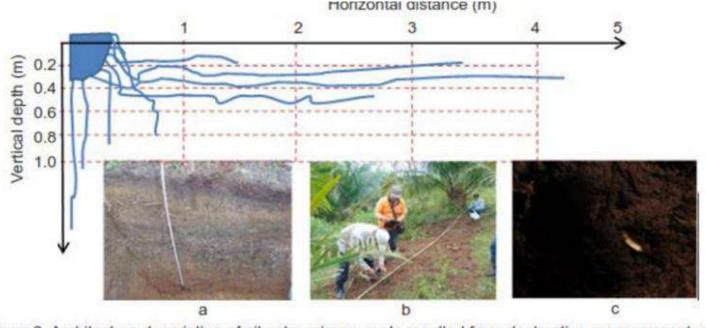


Figure 3 Architecture description of oil palm primary roots resulted from destructive measurement: a Excavation of soil around the palm roots; b Activity of measuring the root length of the palm at ground level; c The primary root tip that grows actively in the soil.

Sanitation will buy you time!

Standard Operating Procedure (SOP): Replanting

Sanitation (removal or debolling) of diseased palm and old palm stump

Preventive treatment (Biological control product)









Standard Operating Procedure (SOP): Replanting

Sanitation (removal or debolling) of diseased palm and old palm stump

Removing the inocula pressure

- Selected sanitation if disease incidences less than 10%.
- Full sanitation if disease incidence more than 10%.
- How about deep ploughing and pulverising??



Standard Operating Procedure (SOP): Replanting

Removing the inocula pressure







Fabricated ripper on D6 bulldozer ploughing through the field, exposing primary roots and old root bole onto soil surface

- 1 metre deep ploughing
- 5 8% of disease incidence after 15 years.



Harrow implement modified for combing the surface roots



Planting hole after 100% deep ploughing field sanitation

Standard Operating Procedure (SOP): Replanting

Preventive treatment (Biological control product)





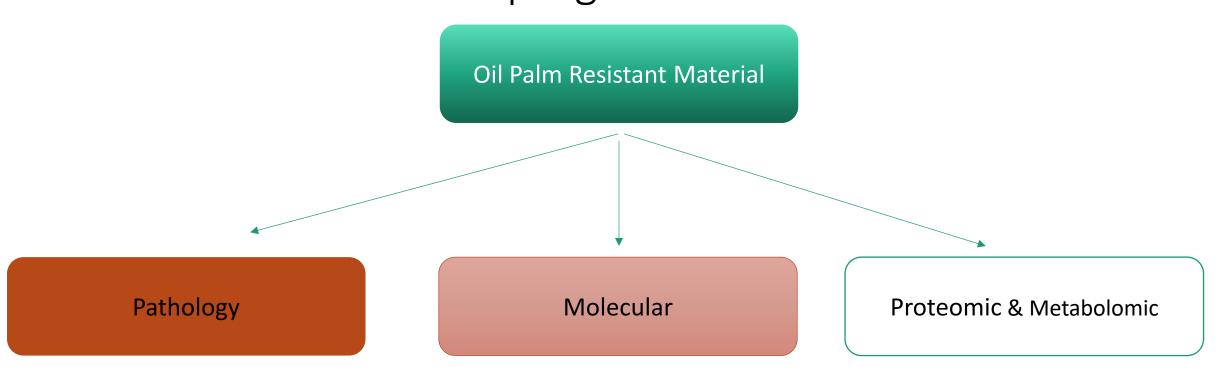




Sebanyak RM 5 juta telah diperuntukkan di dalam Skim Tanam Semula Pekebun Kecil Sawit 2.0 bagi mengaplikasi baja Embio ActinoPLUS dan GanoEF di kawasan berpenyakit.

Syarikat	Pascal	All	IBG	True	
	Biotech	Cosmos	Manufacturing	Trichoderma	
Nama	Embio	GanoEF	IM-Bioguard	Trichoshield	
Produk	ActinoPLUS				
Durasi	10 tahun	10 tahun	2 tahun	3 tahun	
Kejadian					
Penyakit	0.19%	1.32%	1.32% 6.25%		
(10 ha)					





MPOB have started the screening programme since 2016

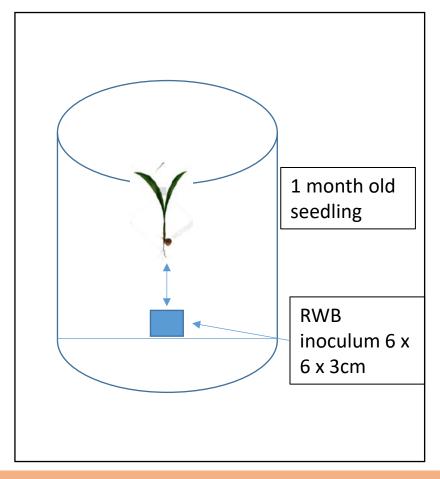
Num.	Progeny Background			
1.	ZRE-Zaire			
2.	CMR-Cameroon			
3.	NGA-Nigeria			
4.	YG-Yangambi			
5.	AV-AVROS			
6.	E-Elmina Dura			
7.	JL-Johor Labis Dura			
8.	BD-Banting Dura			
9.	UR-Ulu Remis Dura			
10.	SD-Serdang Dura			
11.	CD-Chemara Dura			



- Only screen commercial progenies.
- Male x Female backgrounds
- Multiple testing on potential progenies e.g. partial –resistance and susceptible

RESULTS (2ND GENERATION)

ВАТСН	NUM. OF PROGENIES	STATUS
1	14	COMPLETED
2	9	COMPLETED
3	8	COMPLETED
4	12	COMPLETED
5	7	COMPLETED
6	20	COMPLETED
7	23	COMPLETED
8	15	COMPLETED
9	18	COMPLETED
10	22	COMPLETED
11	25	COMPLETED
TOTAL	173	



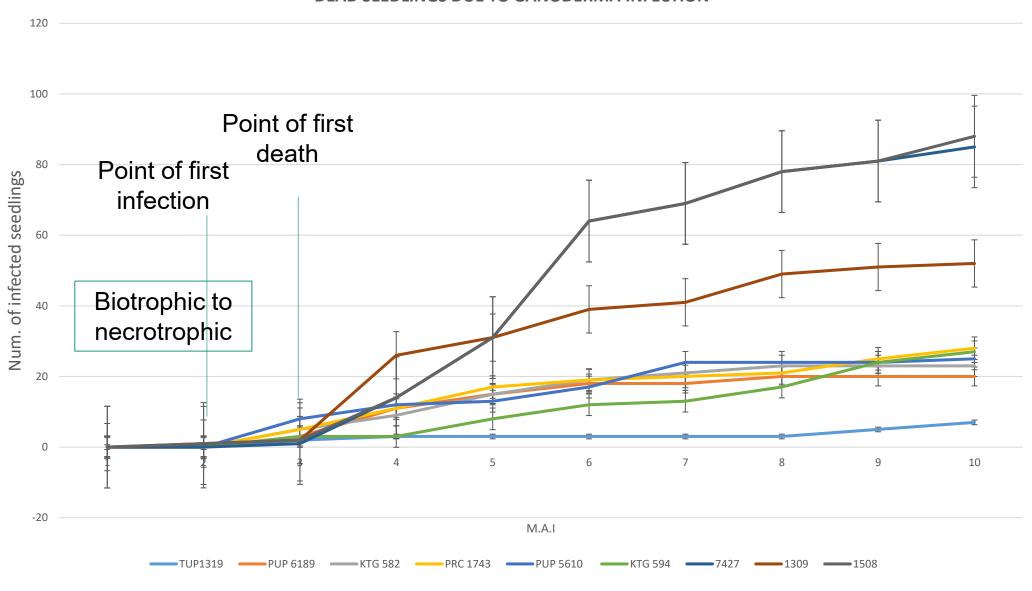
Artifical inoculation used was sitting technique using 1 month old seedling (harmonize with Asian Agri, Indonesia). *Ganoderma* isolate – PER 71

A few interesting pedigree were observed as partial resistance materials. The potential progenies are now undergoing further testing in the nursery and have been planted in *Ganoderma* hot spot areas.

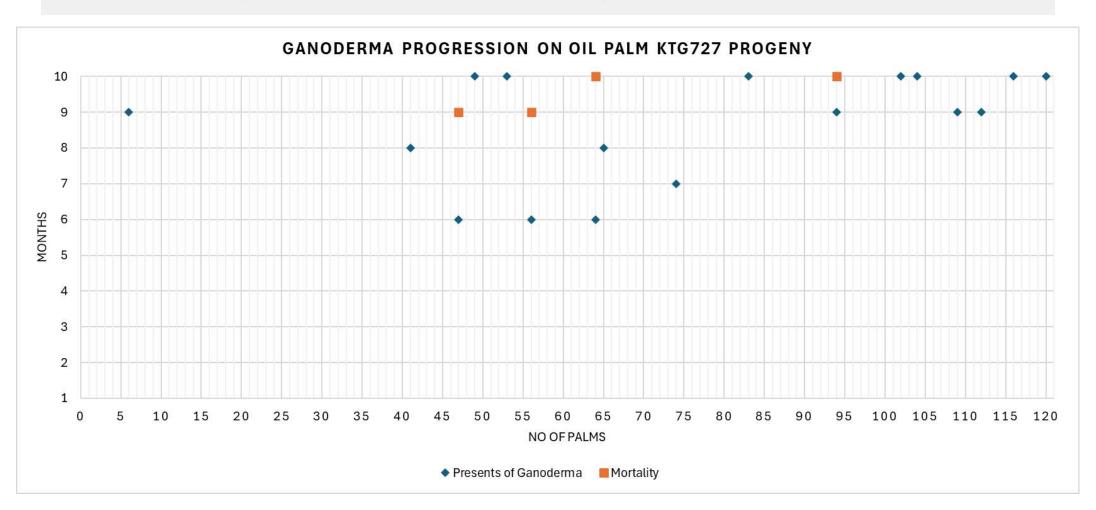
Cross Code	Batch	Cross Type	Background	Number of Samples Screened	% of dead seedlings	Total N Sample	N Repeat	Average Mortality %
KTG 586	6	DxP	CD x (ZRExAV)	100	32.0			
KTG 586	7	DxP	CD x (ZRExAV)	100	47.0	300	3	36.0
KTG 732	11	DxP	CDx(ZRExAV)	100	29.0			
KTG 576	7	DxP	CD x (CMRxAV)	100	47.0	200	2	25.5
KTG 718	11	DxP	CDx(CMRxAV)	100	24.0	200		35.5
KTG 594	6	DxP	CD x (ZRExAV)	100	27.0		3	34.7
KTG 594	7	DxP	CD x (ZRExAV)	100	48.0	300		
KTG 753	11	DxP	CDx(ZRExAV)	100	29.0			
KTG 590	6	DxP	CD x (CMRxAV)	100	30.0			
KTG 590	7	DxP	CD x (CMRxAV)	100	54.0	300	3	36.3
KTG 759	11	DxP	CDx(CMRxAV)	100	25.0			
KTG 574	6	DxP	CD x (ZRExAV)	100	29.0	200	2	
KTG 726	11	DxP	CDx(ZRExAV)	100	24.0			

No.	Cross Code	Batch	Cross Type	Background	Number of Samples Screened	% of dead seedlings	Total N Sample	N Repeat	Average Mortality %
6	KTG 594	6	DxP	CD x (ZRExAV)	100	27.0			
0	KTG 594	7	DxP	CD x (ZRExAV)	100	48.0	300	3	34.7
	KTG 753	11	DxP	CDx(ZRExAV)	100	29.0			
7	KTG 590	6	DxP	CD x (CMRxAV)	100	30.0			
/	KTG 590	7	DxP	CD x (CMRxAV)	100	54.0	300	3	36.3
	KTG 759	11	DxP	CDx(CMRxAV)	100	25.0			
8	KTG 612	8	DxP	CD x (CMRxAV)	120	44.2	220	2	22.1
0	KTG 752	11	DxP	CDx(CMRxAV)	100	22.0	220	2	33.1
9	KTG 609	8	DxP	CD x (ZRExAV)	120	55.8	220	2	34.4
9	KTG 720	11	DxP	Cdx(ZRExAV)	100	13.0	220		
10	KTG 574	6	DxP	CD x (ZRExAV)	100	29.0	200	2	26.5
10	KTG 726	11	DxP	CDx(ZRExAV)	100	24.0	200	2	20.5
11	KTG 568	7	DxP	CD x (CMRxAV)	100	47.0	200	2	25.5
11	KTG 727	11	DxP	Cdx(CMRxAV)	100	4.0	200 2		25.5

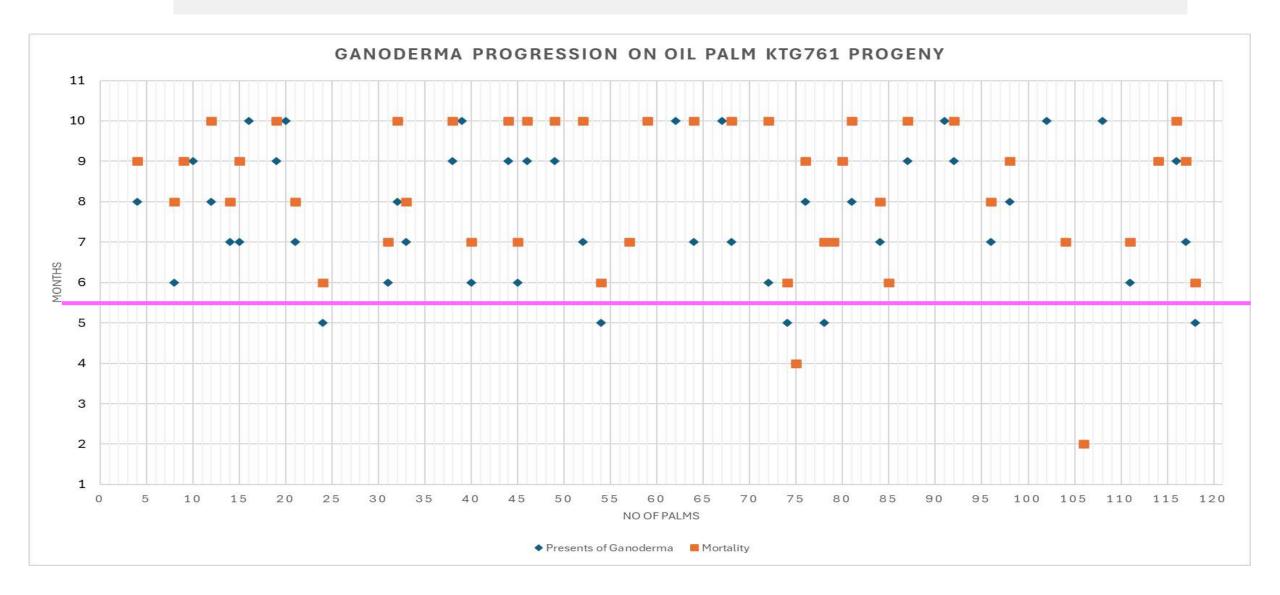
DEAD SEEDLINGS DUE TO GANODERMA INFECTION



POTENTIAL PARTIAL – RESISTANT PROGENY

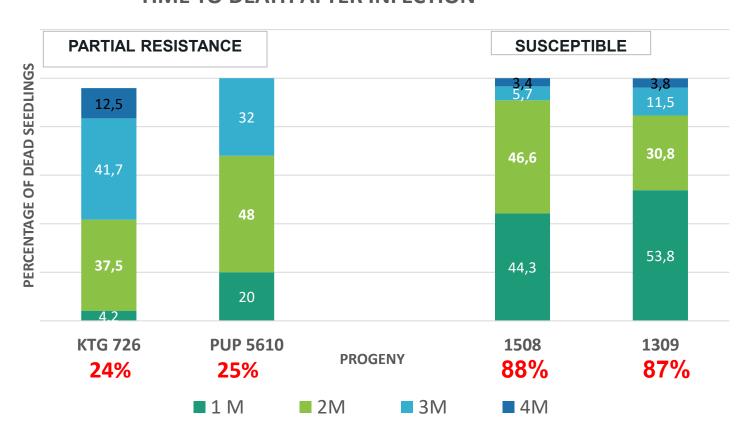


POTENTIAL SUSCEPTIBLE PROGENY



PARTIAL RESISTANCE / SUSCEPTIBLE INFECTION PATTERN

TIME TO DEATH AFTER INFECTION



NOTE:

❖ Partial − resistance progenies: 10 − 15% infected seedlings continue to live on throughout the trial duration

- MPOB collaborates with commercial partner in Indonesia whereby potential partial resistance and susceptible progenies were sent for testing with their *Ganoderma* most aggressive isolate.
- The results patterns was similar albeit higher infections were observed with Indonesia aggressive isolate.

	% of dead seedling due to Ga	lling due to Ganoderma infection			
PROGENY	GB AGGRESSIVE	PER71			
TUP 1269	64	40			
TUP 1299	69	48			
TUP 1310	61	48			
TUP 1249	72	60			
TUP 1273	72	63			
PKC 9211	69	65			
TUP 1260	70	67			
TUP 1217	65	75			
PKC 9197	66	82			
Control Tolerant	19				
Control Susceptible	80				



Moment of Dying (MoD)					Dead or A			
36K				-644				36K
K3 LOD	Method	SNP	Pos	EG11 _Chr	Pos	SNP	Method	K3 LOC
				4	4 COE 4C1	Aff., 1200000200	mrMLM	4.3
				1	4,695,461	Affx-1289698206	pLARmEB	4.6
4.6	pLARmEB	Affx-1289694142	OE 270 214					
3.5	pKWmEB	AIIX-1209094142	95,270,314					
3.3	pKWmEB mrMLM	Affx-952087898	112,529,276	2				
					118,271,241	Affx-952095051	FASTmrMLM	5.3
					122,970,803	Affx-952099729	pLARmEB	7.2
					24,895,668	Affx-1289685211	mrMLM	3.2
					24,893,008	A11X-1289083211	pKWmEB	
					103,897,617	Affx-1289700101	pLARmEB	4.3
				3	118,573,412	Affx-257679660	pLARmEB	3.2
				J	110,575,412	7411X 237 07 3000	mrMLM	
					120,339,768	Affx-1289698543	pLARmEB	3.7
							pKWmEB	3.8
							mrMLM	
	==			4	126,414,425	Affx-1290516698	pKWmEB	3.6
4.4	pKWmEB	Affx-1111256132	4,887,983	5				
3.4	pLARmEB	Affx-952513234	128,796,719					
5.0	mrMLM	Affx-1111380230	28,307,362					
2.5	pLARmEB			8				
3.5	mrMLM	Affx-1291292135	129,211,442					
	pLARmEB			9	94,155,435	Affx-1289703892	mrMIM	4.6
					54,155,455		pKWmEB	3.8
				10	36,655,882	Affx-952467473	pLARmEB	3.0
							mrMLM	4.4
				11	24,378,928	Affx-1295098379	pLARmEB	
				12	489,851	Affx-1292620776		3.6
4.4	pKWmEB			13	,			
	pLARmEB	Affx-1289686295	58,104,994					
				15	53,195,171	Affx-1289709354	pLARmEB	5.4
				16	22 662 154	Affy 1200711752	mrMLM	5.2
				16	23,662,154	Affx-1289711753	pLARmEB	

Resistance related genes

Disease resistance protein RGA4

GATA Transcription Factor 19 (abiotic stress resistance) Phosphomannomutase (immune defenses)

Nudix hydrolase homolog 8 (immune responses)

Elongator complex protein 6 (resistance against pathogens)

Glycine-Rich RNA-Binding Protein 2 RBG2 (stress responses) Probable serine/threonine-protein kinase PBL8 (defense signaling)

YP3-12/2022: RESISTANCE-ASSOCIATED PROTEIN AND METABOLITE PROFILING OF Ganoderma boninense INFECTION IN OIL PALM

Sub-project A: Protein profiling of resistant and susceptible oil palms during *G. boninense* infection Sub-project B: Metabolite profiling of resistant and susceptible oil palms against *G. boninense* infection

Significant Achievements

- Three potential partial resistant and susceptible oil palm seedlings were successfully inoculated with *G. boninense* PER71 and ET61.
- Leaf and root tissues were sampling at 5 time points (Week 0, 1, 2, 4 and 12) after inoculation.
- Preliminary observation: *G. boninense* mycelia were observed at roots of Week 4 and Week 12 (Figs. 1 and 2) with no foliar symptom.
- Protein and RNA were co-extracted from leaf and root tissues (Status 30% completion).
- Preliminary analysis: Differentiation of root and leaf proteome using principal component analysis (PCA).

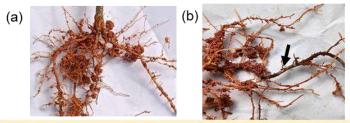


Fig. 1: Representative images of (a) *G. boninense* PER71 and (b) ET61 mycelia at root of Week 4 after inoculation

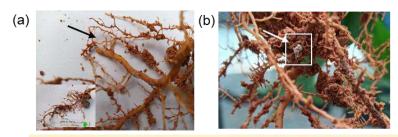
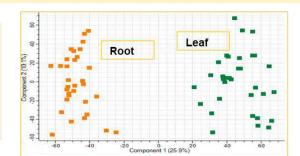


Fig. 2: Representative images of (a) *G. boninense* PER71 and (b) ET61 mycelia at root of Week 12 after inoculation

Fig. 3: Good protein resolution and increased protein identification were observed using the principal component analysis.



Field evaluation is on going – No infection yet after 20 months



CONCLUSION



- Efforts to control must be done continuously control and preventive treaments. Integrated management is essential:
 - I. Early disease detection (diagnostic tools, census).
 - II. Proper sanitation & replanting strategies.
 - III. Fungicide and biological control application.
 - IV. Resistant/tolerant planting materials.
- Fundamental studies improved understanding of infection mechanisms, pathogen diversity, and environmental factors.

CONCLUSION



- SOPs provide practical field guidance for plantations and smallholders.
- Resistant material screening and molecular studies show promise for long-term solutions.
- Continuous R&D, stakeholder collaboration, and adaptive strategies are key to sustaining yield and industry resilience.



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